ESTIMATING CHAOS AND COMPLEX DYNAMICS IN AN INSECT POPULATION

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Abstract. A defining hypothesis of theoretical ecology during the past century has been that population fluctuations might largely be explained by relatively low-dimensional, nonlinear ecological interactions, provided such interactions could be correctly identified and modeled. The realization in recent decades that such nonlinear interactions might result in chaos and other exotic dynamic behaviors has been exciting but tantalizing, in that attributing the fluctuations of a particular real population to the complex dynamics of a particular mathematical model has proved to be an elusive goal. We experimentally tested a model-predicted sequence of transitions (bifurcations) in the dynamic behavior of a population from stable equilibria to quasiperiodic and periodic cycles to chaos to three-cycles using cultures of the flour beetle Tribolium. The predictions arose from a system of difference equations (the LPA model) describing the nonlinear life-stage interactions, predominantly cannibalism. We built a stochastic version of the model incorporating demographic variability and obtained conditional least-squares estimates for the model parameters. We generated 2000 “bootstrapped data sets” with a time-series bootstrap technique, and for each set we reestimated the model parameters. The resulting 2000 bootstrapped parameter vectors were used to obtain confidence intervals for the model parameters and estimated distributions of the Liapunov exponents for the deterministic portion (the skeleton) of the model as well as for the full stochastic model. Frequency distributions of estimated dynamic behaviors of the skeleton at each experimental treatment were produced. For one treatment, over 83% of the bootstrapped parameter estimates corresponded to chaotic attractors, and the remainder of the estimates yielded high-period cycles. The low-dimensional skeleton accounted for at least 90% of the variability in the population abundances and accurately described the responses of populations to experimental demographic manipulations, including treatments for which the predicted dynamic behavior was chaos. Demographic stochasticity described the remaining noise quite well. We conclude that the fluctuations of experimental flour beetle populations are explained largely by known nonlinear forces involving cannibalistic-stage interactions. Claims of dynamic behavior such as periodic cycles or chaos must be accompanied by a consideration of the reliability of the estimated parameters and a realization that the population fluctuations are a blend of deterministic forces and stochastic events.

Key words: bifurcations in dynamic behavior; bootstrap parameter estimates; chaos; complex dynamics; Liapunov exponents; mathematical modeling; periodic cycles; population dynamics, chaotic; population dynamics, nonlinear; population model parameters, estimating; quasiperiodicity; Tribolium.

INTRODUCTION


One particular aspect of nonlinear dynamic theory—
We view such a predicted sequence of transitions as a hypothesis subject to experimental scrutiny in the laboratory. The idea for a “transitions experiment” is straightforward: manipulate biological parameters (such as the birthrate) in order to document experimentally the model-predicted transitions among dynamic states.

There is a demanding prerequisite for both positing and testing a transitions hypothesis: the identification of an adequate mathematical model. Our work began with the derivation of a biologically based stage-structured model (the larva–pupa–adult or “LPA” model) for the flour beetle Tribolium castaneum Herbst (Dennis et al. 1995). We applied this model to the data of Desharnais and Costantino (1980) and, with diagnostic analyses of time-series residuals, showed that the LPA model did an excellent job of predicting responses to the experimental perturbations in population stage structure (Dennis et al. 1995). More recently, Benoît et al. (1998) conducted a set of manipulation experiments using the species T. confusum that provided an independent validation of the LPA model.

Following the documentation of the LPA model, we conducted two long-term transitions experiments. In the first transitions experiment, we manipulated the adult mortality rates in beetle cultures and observed transitional changes in the population dynamics corresponding to the bifurcations predicted by the mathematical model (Costantino et al. 1995, Dennis et al. 1997). Maximum-likelihood parameter estimates were obtained, 95% confidence intervals were calculated from profile likelihoods, and the variance–covariance matrices for both the control and treatment replicates were presented. The LPA model did not forecast chaotic attractors in this experiment, and none were observed experimentally (Rohani and Miramontes 1996, Dennis et al. 1997). The model-predicted transitions in population dynamics from equilibria to periodic cycles to aperiodic cycles were documented statistically in the data (Costantino et al. 1995, Dennis et al. 1997).

In this paper we present the results of the second transitions experiment, in which a model-predicted route to chaos was forecast. Our initial report of chaos (Costantino et al. 1997) was based on a point estimate of a set of parameters. What kind of confidence statement can be attached to such a report? Claims of chaos or even stable-point equilibria or stable cycles are typically subject to estimation error (Dennis et al. 1995, Falck et al. 1995a, b). However, it is difficult to produce and interpret confidence statements about dynamic behavior. Some approaches that have been used are profile likelihood methods (Dennis et al. 1995, Bailey et al. 1997), parametric bootstrapping methods (Dennis and Taper 1994), and semiparametric bootstrapping methods (Falck et al. 1995a, b). Of these methods, the semiparametric bootstrapping approach is theoretically more robust to misspecification of the noise distribution in the time-series model.

Our objective here is to explore the robustness of
the results of our second transitions experiment and to ascertain confidence levels for estimates of dynamic behavior. We begin by refitting the LPA model using a noise structure based on demographic stochasticity that describes the data more accurately than our original environmental noise structure (Costantino et al. 1997). We calculate parameter estimates for the stochastic model using the method of conditional least squares (Klimko and Nelson 1978, Tong 1990, Dennis et al. 1995). To calculate confidence intervals, we modify the semiparametric bootstrapping method used by Falk et al. (1995a, b). The estimated deterministic portion of the model is combined with a resampling of the model residual vectors to generate 2000 “bootstrapped data sets.” Model parameter vectors are estimated for each bootstrapped data set, and both deterministic and stochastic Liapunov exponents are computed for each combination of the 2000 bootstrapped parameter vectors and eight experimental treatments. With the 2000 bootstrapped parameter vectors we obtain frequency distributions of Liapunov exponents and frequencies of attractor types predicted by the deterministic portion of the model for each experimental treatment. We show that the flour beetle populations display an array of complex dynamic behaviors that includes a strong influence of chaos. We discuss the difference between the deterministic and the stochastic Liapunov exponents, and conclude that the stochastic Liapunov exponents probably should not be used as a benchmark of chaos. We caution that claims of “dynamic behavior” such as periodic cycles and chaos must be accompanied by a consideration of the reliability of the estimated parameters and a realization that the population fluctuations are a combination of the influence of the underlying deterministic dynamics as well as of stochastic events. We discuss the interplay of stochastic and deterministic forces (Bartlett 1990, Renshaw 1994), the meaning of chaos in a stochastic system, and the importance of incorporating stochastic forces along with studying and quantifying the influence of underlying deterministic forces in a population model.

METHODS AND DATA

LPA model

Many species of Tribolium are cannibalistic (Park et al. 1965), including the species Tribolium castaneum (Herbst) used in this project. The LPA (larva–pupa–adult) model is built on the assumption that the nonlinear interactions among the life-cycle stages caused by cannibalism are the driving mechanisms of the dynamics for this species. The LPA model is a system of three difference equations that relate the numbers of animals at time t to the number of animals at time t − 1:

\[ L_t = bA_{t-1} \exp(-c_aA_{t-1} - c_pA_{t-1}) \]  

\[ P_t = L_{t-1}(1 - \mu_a) \]  

\[ A_t = P_{r-1} \exp(-c_pA_{r-1}) + A_{r-1}(1 - \mu_a). \]

In this model formulation (Dennis et al. 1995, 1997), \( L_t \) is the number of feeding larvae (referred to as the “L-stage”), \( P_t \) is the number of large larvae, non-feeding larvae, pupae, and callow adults (called the “P-stage”), and \( A_t \) is the number of sexually mature adults (“A-stage animals”) at time t. The unit of time is 2 wk and is, approximately, the average amount of time spent in the feeding larval stage under our experimental conditions. The unit of time is also approximately the average duration of the P-stage. The quantity \( b \) is the number of larval recruits per adult per unit of time in the absence of cannibalism. The fractions \( \mu_l \) and \( \mu_a \) are the larval (l) and adult (a) rates of mortality, respectively, in one time unit. The exponential nonlinearities account for the cannibalism of eggs (e) by both larvae and adults and the cannibalism of pupae by adults. The fractions \( \exp(-c_aL_{t-1}) \) and \( \exp(-c_pA_{t-1}) \) are the probabilities that an egg is not eaten in the presence of \( L_{t-1} \) larvae and \( A_{t-1} \) adults, respectively, in one time unit. The fraction \( \exp(-c_pA_{t-1}) \) is the survival probability of a pupa (p) in the presence of \( A_{t-1} \) adults in one time unit. Eqs. 1–3 represent the deterministic population forces upon which we build our stochastic model.

Demographic variability

Different types of stochastic mechanisms produce different patterns of variability. In particular, two broad classes of stochastic mechanisms important to populations have been widely discussed: demographic stochasticity and environmental stochasticity (May 1974b, Shaffer 1981). In order to connect a deterministic population model with time-series data, the population model has to be converted into a stochastic model (Dennis et al. 1995). The types of stochastic mechanisms affecting the population therefore have to be carefully considered and formulated.

“Demographic stochasticity” is the variability in population size caused by independent random contributions of births, deaths, and migrations by individual population members. Demographic stochasticity can be modeled by taking each individual’s net contribution to the population in a unit of time to be the outcome of an independent, discrete random variable. A simple example is survival: an individual contributes “0” to the population in the next time unit with probability \( \mu \), where \( \mu \) is the probability of death during the time unit, and “1” to the population with probability \( 1 - \mu \). If many individuals are members of a homogeneous cohort, then demographic events can often be aggregated analytically at the cohort level into a known probability distribution for cohort size at the next time unit.

For instance, suppose each member of a cohort of \( n_{r-1} \) individuals at time \( t - 1 \) has the same unit-time survival probability, \( 1 - \mu \). A deterministic model for the number of survivors at time t is \( n_t = n_{r-1}(1 - \mu) \). If each cohort member were instead subjected independently to a simple, random survival process, the cohort at time t would be a random variable, \( N_t \), with
a binomial\(\binom{n - 1, (1 - \mu)}{l_{t - 1}}\) distribution. Note that the binomial distribution for \(N_t\) is conditional on the attained value, \(N_{t-1} = n_{t-1}\), of the previous cohort size. The conditional mean of \(N_t\) is \(n_{t-1}(1 - \mu)\) (the deterministic expression), and the conditional variance of \(N_t\) is \(n_{t-1}(1 - \mu)\mu\) (that is, the conditional variance is proportional to \(n_{t-1}\)).

We constructed a stochastic version of the LPA model containing demographic variability. In the stochastic version, we used the binomial and Poisson distributions to describe the aggregation of demographic events within the life stages. The simplest case is the P-stage model (Eq. 2). In the stochastic model, \(P_t\) is a random variable. The distribution of \(P_t\) given \(L_{t-1} = l_{t-1}\) was taken to have a binomial\(\binom{l_{t-1}, (1 - \mu)}{l_{t-1}}\) distribution. The A-stage (Eq. 3) is the sum of two survival processes: recruits from the P-stage, denoted \(R_t\), and surviving adults, denoted \(S_t\). We assumed that \(R_t\) given \(P_{t-1} = p_{t-1}\) and \(A_{t-1} = a_{t-1}\) has a binomial\(\binom{p_{t-1}, \exp(-c_{sa_{t-1}})}{1}\) distribution; the P-stage survival probability is the nonlinear function of \(a_{t-1}\) from Eq. 3. Also, \(S_{t-1}\) was assumed to have a binomial\(\binom{a_{t-1}, (1 - \mu)}{1}\) distribution.

The L-stage is a compound process: a random number of potential recruits is produced (with conditional mean \(ba_{t-1}\)), and each potential recruit subsequently undergoes a survival process in which the conditional survival probability, \(\exp(-c_{da_{t-1}} - c_{ca_{t-1}})\), depends on the system state variables \(l_{t-1}\) and \(a_{t-1}\) (Eq. 1). We assumed that the number of potential recruits has a Poisson\(\binom{ba_{t-1}}{1}\) distribution (i.e., a Poisson distribution with mean \(ba_{t-1}\)), and that the number of subsequent survivors has a binomial distribution. An elementary property is that if a random variable \(Y\) has a binomial\(\binom{n, p}{1}\) distribution given \(N = n\), and \(N\) has a Poisson\(\lambda\) distribution, then the resulting distribution for \(Y\) is Poisson\(\lambda p\) (Boswell et al. 1979). Thus, the conditional distribution of \(L_t\) given \(L_{t-1} = l_{t-1}\) and \(A_{t-1} = a_{t-1}\) becomes a Poisson distribution with mean \(ba_{t-1}\exp(-c_{da_{t-1}} - c_{sa_{t-1}})\).

Our stochastic demographic LPA model thus takes the conditional one-time-step distributions of \(L_t\), \(P_t\), and \(A_t\) to be independent, discrete probability distributions:

\[
L_t \sim \text{Poisson}(ba_{t-1}\exp(-c_{da_{t-1}} - c_{sa_{t-1}})) \quad (4)
\]

\[
P_t \sim \text{binomial}(l_{t-1}, (1 - \mu)) \quad (5)
\]

\[
R_t \sim \text{binomial}(p_{t-1}, \exp(-c_{sa_{t-1}})) \quad (6)
\]

\[
S_t \sim \text{binomial}(a_{t-1}, (1 - \mu)) \quad (7)
\]

\[
A_t = R_t + S_t \quad (8)
\]

Here ‘\(\sim\)’ means ‘is distributed as.’

For most statistical inferences reported in this paper, we undertook an additional modification of the stochastic demographic LPA model (Eqs. 4–8). We transformed the observations so that the stochastic demographic model would be approximated by a nonlinear autoregressive (NLAR) model. An NLAR model has the form

\[
X_t = h(X_{t-1}) + E_t \quad (9)
\]

where \(X_t\) is a state variable at time \(t\), the function \(h(\cdot)\) is the ‘skeleton’ (underlying deterministic equation for \(X_t\); Tong 1990), \(E_t\) is a normally distributed random variable with mean zero and variance \(\sigma^2\), and \(E_t, E_{t-1}, \ldots\) are uncorrelated. NLAR models have some advantages and conveniences for our analyses. First, parameter estimates retain a theoretical robustness to departures of the data from distributional assumptions (see Parameter estimation, below). Second, the numerically intensive calculations and simulations that we needed involve straightforward algorithms for least squares and normal random-variable generation. Finally, NLAR model evaluation can be based on familiar diagnostic methods using residuals (see Dennis et al. 1995).

The idea behind the NLAR approximation is to find a transformation, \(X_t = g(N_t)\), where \(N_t\) is a state variable in the original model, for which the distribution of \(X_t\) given \(X_{t-1} = x_{t-1}\) is approximately normal with a constant variance (that is, a variance that does not depend on \(x_{t-1}\)). If a variable \(N_t\) has a Poisson distribution with mean \(\lambda\), then a well-known result from statistics is that \(X_t = \sqrt{N_t}\) has approximately a normal distribution with a mean of \(\sqrt{\lambda}\) and a constant variance that does not depend on the value of \(\lambda\) (Rao 1973). Thus, the square-root transformation stabilizes and normalizes the variance for the L-stage variable (Eq. 4). Also, the binomial distributions in the P-stage and A-stage models (Eqs. 5–8) are well approximated by Poisson distributions (e.g., Rice 1995), and so the square-root transformation approximately stabilizes and normalizes those variables as well. Thus, a demographic NLAR approximation for a variable \(X_t\) is built by transforming the deterministic equation for \(N_t\) to the square-root scale, producing the transformed map given by \(X_t = h(X_{t-1})\). Adding normal noise for stochasticity produces the NLAR model (Eq. 9). The NLAR model, in conjunction with the square-root transformation, can be regarded as a general method of incorporating demographic stochasticity into a deterministic model. We note that an NLAR process using a logarithmic instead of a square-root transformation is a standard way of incorporating environmental variability (Dennis and Taper 1994, Dennis et al. 1995, Costantino et al. 1997).

The LPA model (Eqs. 1–3) has three state variables, and the demographic NLAR approximation applied to the LPA model produces a multivariate NLAR process. In Eq. 9, \(X_t\) becomes a vector of transformed state variables, \(h(\cdot)\) becomes a vector-valued NLAR process, and \(E_t\) becomes a vector of noise variables. Applying the square-root transformation to Eqs. 1–3, and adding noise, yields the following for the stochastic demographic LPA model:
\[ \sqrt{E} = \sqrt{bA_{j-1} \exp(-c_{ij}A_{i-1}) - c_{ij}A_{j-1}} + E_{i}, \quad (10) \]
\[ \sqrt{P} = \sqrt{E_{j-1}(1 - \mu_{j}) + E_{2j}} \quad (11) \]
\[ \sqrt{A} = \sqrt{P_{j-1} \exp(-c_{ij}A_{j-1}) + A_{j-1}(1 - \mu_{j}) + E_{3j}}. \quad (12) \]

The terms \( E_i = (E_{1i}, E_{2i}, E_{3i}) \) constitute a random noise vector assumed to have a joint normal probability distribution with a mean vector of zeros and a diagonal variance–covariance matrix denoted by \( \Sigma \). The demographic nature of the stochasticity would stipulate that the noise variables are uncorrelated with each other within a time unit (off-diagonal elements of the matrix \( \Sigma \) are zero) as well as uncorrelated through time. The deterministic skeleton of the model (Eqs. 1–3 transformed) is obtained by setting \( \Sigma = 0 \), or equivalently, by letting \( E_{1i}, E_{2i}, \text{and } E_{3i} \) equal zero.

As written, the demographic NLAR model with normal noise (Eqs. 10–12) can produce negative values for the (transformed) state variables, because the normal noise variables have a nonzero chance of being negative and of large magnitude. Such instances have negligible probabilities under the model parameter values associated with our data. In the rare instances in which noise produced a negative value of a state variable, the state variable was set to zero. Unlike the situation for the logarithmic transform under environmental variability, zero values for state variables are used in the square-root transform under demographic variability just like any other values and do not require special handling.

For the data from the second transitions experiment (Costantino et al. 1997), we compared the demographic (square-root) and environmental (logarithmic) transformations, as well as a third transformation involving a weighted mixture of demographic and environmental stochasticity. We estimated the parameters of the resulting stochastic LPA models and conducted diagnostic analyses of the residuals (homoscedasticity, autocorrelations, and normality of residuals). Our comparative diagnostic analyses, to be reported in a separate paper, suggested that the model for demographic stochasticity is superior. This is not surprising. The second transitions experiment produced time series with many low (single-digit) values of state variables, due to the experimental manipulations of the treatment populations. Demographic variability is known to be more important for low population sizes (see Simberloff 1988). Therefore, despite the fact that we used environmental noise in our first report (Costantino et al. 1997), the analyses in the present paper are based on the stochastic demographic LPA model given by Eqs. 10–12. However, the qualitative predictions of the fitted models (cycles, chaos, etc.) for both formulations were nearly identical.

The dominant Liapunov exponent

The dominant Liapunov exponent (LE) quantifies the tendency of a deterministic model to be sensitive to initial conditions. It is an index calculated for a trajectory of a deterministic model. At each point of a trajectory there is a maximum rate at which all nearby trajectories can diverge from or converge to one another. The LE is the average of these local maximum rates taken over all points of the trajectory. Thus, in this sense the LE of a trajectory can be thought of as a long-run average rate of exponential divergence or convergence of nearby trajectories. For almost all trajectories converging to an attractor such as a stable point, stable cycle, or strange attractor, the LEs of the trajectories will be identical to the LE of the attractor because the effects of the initial conditions on the averages become washed out. The LE of an attractor is defined as the LE of a dense trajectory on the attractor (that is, a trajectory that comes arbitrarily close to every point on the attractor). A nonchaotic trajectory such as a periodic cycle can exist on a chaotic attractor, but such a trajectory is not dense.

For a bounded trajectory that is not asymptotic to a periodic cycle, a positive LE is an indication of a chaotic attractor (Alligood et al. 1997). If the LE is positive, tiny changes in initial conditions are magnified on average along the trajectory. A negative LE indicates that trajectories differing by tiny changes in initial conditions tend on average to converge; such is the case with a stable point equilibrium or a stable periodic cycle. A stable invariant loop has an LE of zero: on average along a trajectory tiny changes in initial conditions are neither magnified nor diminished.

The LE for the trajectories of the LPA model (Eqs. 1–3) can be computed by iteration. Let \( J \) be the Jacobian matrix of the LPA model evaluated at the values \( L_0, P_0, \text{and } A_0 \) that is,

\[ J = \begin{pmatrix} -c_{ij}bA_{ij}1 & b(1 - c_{ij}A_i)g_{1j} & 0 \\ -c_{ij}bA_{ij}1 & 0 & 0 \\ -c_{ij}bA_{ij}1 & -c_{ij}P_{ij}g_{2j} + (1 - \mu_{ij}) & 0 \end{pmatrix} \quad (13) \]

where, for brevity, \( g_{1j} = \exp(-c_{ij}L_i - c_{ij}A_i) \) and \( g_{2j} = \exp(-c_{ij}A_i) \). Also, let \( \lambda_i \) be defined as a “geometric mean” of the norm of the products of the Jacobian matrices:

\[ \lambda_i = (1/t)\ln \left| JJ_{i+1}J \cdots J_m \right|. \quad (14) \]

Here \( \left| \right| \) denotes any matrix norm (we used the Euclidean norm, \( |M| = \sqrt{\sum \sum \left| m_{ij} \right|^2} \), where \( m_{ij} \) is the element in the \( i \)th row and \( j \)th column of a matrix \( M \)). The LE can in principle be computed as

\[ \lambda = \lim_{t \to \infty} \lambda_i \quad (15) \]

for any positive initial condition. This definition is analogous to the eigenvalue for linear stability analysis; here the linearization is “averaged” over the long-term trajectory of the population as it visits points on the attractor. If the attractor is a stable point equilibrium, then the LE is identical to the logarithm of the modulus.
of) the largest eigenvalue of $J_t$ evaluated at the equilibrium (the eigenvalue commonly used in stability analysis of a discrete-time system; May 1974b).

The definition of the LE can be extended to the stochastic model (Eqs. 10–12). In this case, the average used to calculate the trajectory LE does not converge to the LE averaged on a deterministic attractor. Rather, the trajectory LE converges to the LE averaged over the stationary probability distribution for the stochastic population (Crutchfield et al. 1982, Kifer 1986, McCaffrey et al. 1992, Ellner and Turchin 1995). However, one must be careful about the interpretation of the “sensitivity to initial conditions” implied by a positive stochastic Liapunov exponent (SLE), since stochastic models do not have chaotic attractors and two independent stochastic processes will always drift apart due to the random nature of their dynamics. A positive SLE is an indication that a population spends a large amount of time in a state space region where deterministic trajectories would diverge. Such regions include not only strange attractors but also regions near unstable equilibria and other repellers; in stochastic models, such regions can be revisited often. Thus, the SLE measures something different from the deterministic LE. We develop this point further in the Discussion, below.

There are several practical problems with using Eq. 15 directly to compute $\lambda$. The matrix product is numerically unstable for large $t$, and, unfortunately, extremely large values of $t$ are usually needed for convergence. Care must be taken to minimize the accumulation of numerical roundoff error. Finally, intelligent decisions must be made about the criterion for convergence, especially when $t$ is large. In the Appendix we describe our algorithm for overcoming these difficulties.

**Experimental data**

The experiment, described below, was designed explicitly to test the predictions of the deterministic skeleton of the LPA model. Parameter estimates from an earlier experiment (Costantino et al. 1995) predicted a sequence of changes in dynamical behavior from equilibria to invariant loops and periodic cycles to chaos to three cycles as the parameter $c_m$ was increased from 0 to 1 with $\mu_5 = 0.96$ (Dennis et al. 1997). In the current study we test that prediction.

We experimentally set the adult mortality rate at $\mu_5 = 0.96$ and manipulated the adult recruitment rate so that it would equal $P_a \exp(-c_m A)$, with values of $c_m$ set at 0.00, 0.05, 0.10, 0.25, 0.35, 0.50, and 1.00. There was also an unmanipulated control treatment. Twenty-four cultures of the RR strain of the flour beetle *Tribolium castaneum* were initiated with 250 L-stage insects, 5 P-stage animals, and 100 A-stage sexually mature young adults. Three populations were randomly assigned to each of the eight treatments. Each population was maintained in a half-pint (237 mL) milk bottle with 20 g of standard media and kept in a dark incubator at 32°C. Every 2 wk the L-, P-, and A-stages were censused and returned to fresh media, and dead adults were counted and removed. This procedure was continued for 80 wk. Adult mortality was set by removing or adding adults at the time of a census to make the fraction of adults that died during the interval equal to 0.96. Recruitment rates into the adult stage were manipulated by removing or adding young adults at the time of a census to make the number of new adult recruits consistent with the treatment value of $c_m$. To counter the possibility of genetic changes in life-history characteristics, at every other census the adults returned to the population after the census were obtained from separate stock cultures maintained under standard laboratory conditions. Earlier analyses of this experiment were presented by Costantino et al. (1997) and Desharnais et al. (1997).

**Parameter estimation**

The method of conditional least squares (CLS) was used for estimation of the parameters in the stochastic demographic LPA model (Eqs. 10–12). CLS methods relax many distributional assumptions about the noise variables in the vector $E_t$ (Klimko and Nelson 1978, Tong 1990). CLS estimates are consistent (converge to the true parameters as sample size increases), even if $E_t$ is non-normal and autocorrelated, provided the stochastic model (Eq. 9) has a stationary distribution.

CLS estimates are based on the sum of squared differences between the value of a variable observed at time $t$ and its expected (or one-step forecast) value, given the observed state of the system at time $t - 1$. For fitting the LPA model to a single time series, there are three such conditional sums of squares:

$$Q_1(b, c_m, c_a) = \sum_{i=1}^{q} \left( \sqrt{I_i} - \sqrt{I_{i-1}} \exp(-c_a I_{i-1} - c_m a_{i-1}) \right)^2$$

$$Q_2(\mu_5) = \sum_{i=1}^{q} \left( \sqrt{P_i} - \sqrt{P_{i-1} (1 - \mu_5)} \right)^2$$

$$Q_3(c_m) = \sum_{i=1}^{q} \left( \sqrt{a_i} - \sqrt{a_{i-1} \exp(-c_m a_{i-1}) + a_{i-1} (1 - \mu_5)} \right)^2$$

Here $I_t$, $P_t$, and $a_t$, $t = 0, 1, \ldots, q$ are the observed census counts for the three life stages. The conditional sums of squares are constructed on the square-root scale because that is the scale on which we assume noise is additive (Eqs. 10–12). Parameter $\mu_5$ was estimated directly in the control cultures as a binomial probability from the counts of dead adults and the number of adults at risk of mortality at each census interval.
To estimate \( c_p \) in the control cultures, Eq. 18 was used with the value of \( \mu_p \) fixed at the binomial estimate. Eq. 18 was not used for the treatment cultures, because \( c_p \) and \( \mu_p \) were experimentally fixed in those cultures.

CLS estimates for one population minimize the conditional sums of squares (Eqs. 16–18). Our data (see Experimental data, above) consisted of 24 populations: three populations in each of eight treatments, with each population initiated at time \( t = 0 \) and halted at time \( t = 40 \). If \( Q_{ij} \) is the conditional sum of squares for state variable \( i \) in the \( j \)th treatment in the \( k \)th population, then the total conditional sum of squares for the \( i \)th state variable was calculated as

\[
Q_i = \sum_{j=1}^{8} \sum_{k=1}^{3} Q_{ij}.
\]

(19)

Thus, just one set of CLS parameter estimates was obtained for the whole experiment. Three separate numerical minimizations are required, one for each \( Q_i \). We found the Nelder-Mead simplex algorithm (Press et al. 1992) convenient. Alternatively, the CLS estimates can be obtained by minimizing \( Q_1 \), \( Q_2 \), and \( Q_3 \) with a standard nonlinear regression package.

The noise variances (diagonal elements in the variance-covariance matrix \( \Sigma \) of \( E \), Eqs. 10–12) were estimated using the conditional residuals (Dennis et al. 1995). We estimated noise covariances as well, to check the assumption under demographic variability that the variance-covariance matrix \( S \) are not correlated (that is, the off-diagonal elements of \( \Sigma \) are 0). Let \( \mathbf{x} \) denote the vector \([\sqrt{1}, \sqrt{1}, \sqrt{2}, \ldots] \) of observed, transformed stage abundances in a population at time \( t \). Thus, \( \mathbf{x}_0, \mathbf{x}_1, \mathbf{x}_2, \ldots \) is a time series of vector observations arising from one of the experimental populations. Also, let \( \mathbf{h}(\mathbf{x}_{t-1}) \) denote the vector of functions in the model skeleton (Eq. 9) evaluated at \( \mathbf{x}_{t-1} \) using the CLS parameter estimates. Each population in the experiment yielded a series of residual vectors defined as

\[
\hat{\mathbf{e}}_t = \mathbf{x}_t - \mathbf{h}(\mathbf{x}_{t-1})
\]

(20)

for \( t = 1, 2, \ldots, 40 \). The noise variances and covariances for the three control populations were estimated from the sum of squares and crossproducts matrix calculated with the 120 residual vectors. A separate variance-covariance matrix was estimated for the 21 treatment populations, to allow for the possibility that the adult manipulations might alter the stochastic variability in the L and P stages, particularly the variability in L-stage recruitment. The control residual vectors were \( 3 \times 1 \); however, the treatment residual vectors were \( 2 \times 1 \) (L and P stages only) because the experimental manipulations rendered the adult populations deterministic.

The LE and SLE corresponding to each experimental treatment are functions of LPA model parameters. Point estimates of the LE for each treatment were obtained from the deterministic model (Eqs. 1–3) evaluated at the CLS parameter estimates of \( b, c_p, c_{ar} \), and \( \mu_p \), and relevant CLS (control) or fixed (treatment) values of \( c_p \) and \( \mu_p \). Point estimates of the SLE for each treatment were calculated with the stochastic model (Eqs. 10–12), using the CLS estimates of \( b, c_{p}, c_{ar}, \) and \( \mu_p \), relevant values of \( c_p \) and \( \mu_p \), and the estimates of the noise variances obtained from the residuals of the fitted model.

We also calculated maximum-likelihood (ML) estimates for the LPA parameters in the Poisson-binomial model (Eqs. 4–8). The purpose was to compare the CLS estimates from the NLR approach (Eqs. 10–12) with the ML estimates. With the Poisson-binomial model, the likelihood function for each state variable is a product of one-step Poisson or binomial probabilities. For illustration, we focus on the time series emerging from one population. The likelihood function for the parameters in the L-stage model is

\[
L_1(b, c_{aL}, c_{el}) = \prod_{t=1}^{q} \exp[ba_{t-1} \exp(-c_{aL}a_{t-1} - c_{aL}a_{t-1})] \\
\times [ba_{t-1} \exp(-c_{aL}a_{t-1} - c_{aL}a_{t-1})]^{a_{t-1}}.
\]

(21)

This is a product of Poisson probabilities for the values \( l_i(t = 1, 2, \ldots, q) \), with means as indicated by Eq. 4. The likelihood functions for the parameters in the models for the other state variables are products of binomial probabilities:

\[
L_2(\mu_p) = \prod_{t=1}^{q} \left( \frac{l_t-1}{p_t} \right) (1 - \mu_p)^{p_t}(\mu_p)^{l_t-1}.
\]

(22)

\[
L_3(c_{pa}) = \prod_{t=1}^{q} \left( \frac{p_t-1}{r_t} \right) \exp(-c_{pa}a_{t-1})^{r_t} \\
\times [1 - \exp(-c_{pa}a_{t-1})]^{p_t-1}.
\]

(23)

\[
L_4(\mu_a) = \prod_{t=1}^{q} \left( \frac{a_t-1}{s_t} \right) (1 - \mu_a)^{s_t}(\mu_a)^{a_t-1}.
\]

(24)

Here \( s_t \) is the number of adults surviving from time \( t - 1 \) \( (a_{t-1} \) minus the number of dead adults counted at time \( t \) \), and \( r_t = a_t - s_t \). The ML estimates are the values of the unknown parameters \( c_{aL}, c_{ar}, \mu_p \), and in the control cultures \( c_p \) and \( \mu_p \), that jointly maximize the likelihood functions. As with the CLS estimates, all the time series from the experiment were combined to obtain one set of ML parameter estimates. The actual likelihood functions used were products of functions like Eqs. 21–24 taken over all the experimental populations. The ML estimates were calculated by numerical maximization (Eqs. 22 and 24 also admit simple closed-form solutions for the ML estimates).

Model evaluation

In two previous studies we withheld at least half of the data from the parameter estimation process and
used the withheld data in model prediction/validation analyses (Dennis et al. 1995, 1997). In the present study we used all the data in estimation in order to concentrate all the available information into confidence statements about dynamics. The analyses described here are therefore to be interpreted as goodness-of-fit analyses. Nonetheless, because the model has so few parameters, the data are so abundant, the experimental manipulations are so distinct, and the model-predicted responses to those manipulations are so sharp, evidence that the LPA model fits this experiment serves as a fairly severe test of the model.

Our main criterion for evaluating goodness-of-fit was $R^2$. We calculated a generalized $R^2$ statistic to quantify the overall influence of the LPA model skeleton. For state variable $i$ in the $j$th experimental treatment, the conditional sum of squares $Q_{ij}$ was calculated by using the CLS parameter estimates in Eq. 16, 17, or 18. A second conditional sum of squares, $\hat{S}_{ij}$, was calculated using the sample mean of the (square-root transformed) state variable values as the one-step predicted value.

For example, for the L-stage, using subscripts to denote the $k$th population in the $j$th treatment,

$$\hat{S}_{ij} = \sum_{k=1}^{3} \left( \sqrt{j_{ik}} - m_{ij} \right)^2$$

where

$$m_{ij} = \frac{1}{120} \sum_{k=1}^{3} \sum_{t=1}^{40} \sqrt{j_{ik}}$$

is the sample mean of the square-root transformed L-stage abundances in the $j$th treatment. The generalized $R^2$ value for state variable $i$ in the $j$th treatment was then defined as

$$R_{ij}^2 = 1 - \frac{Q_{ij}}{\hat{S}_{ij}}.$$  \hspace{1cm} (25)

An overall $R^2$ was calculated for the $i$th state variable as

$$R_i^2 = 1 - \left( \frac{\sum_{j=1}^{a} Q_{ij}}{\sum_{j=1}^{a} \hat{S}_{ij}} \right).$$

(26)

It will be noted that each treatment retains its own mean in the overall $R^2$ formula, rather than pooling observations across treatments to calculate a common mean.

The model was further evaluated using residual analyses. If the skeleton captures the essential dynamical behavior, the leftover variability on some appropriate scale should be just noise. Additionally, the demographic scale we selected for the noise (as reflected through Eqs. 10–12) should turn out to be an appropriate one. In order to examine these assumptions, the residuals for each stage in each culture (Eq. 20) were subjected to the following diagnostic procedures. First, the first-order and second-order autocorrelations of the residuals were calculated. Autocorrelation of residuals indicates a relationship between successive prediction errors, and thus might suggest a systematic lack of fit between model and data. Second, the residuals were screened in standard diagnostic plots, including normal quantile-quantile plots, and tested for lack of fit to a normal distribution using the Lin-Mudholkar statistic (see Tong 1990). The Lin-Mudholkar statistic was designed for power against asymmetrical alternatives (Lin and Mudholkar 1980). Asymmetry is a main feature that is visually detectable in normal quantile–quantile plots; we report the Lin-Mudholkar statistic in this paper to condense the information from many figures. While CLS estimates are robust to autocorrelation and departure from normality, the diagnostic screening helps determine whether the demographic noise model adequately portrays the stochastic variability in the system.

**Bootstrap confidence intervals**

If the experiment could somehow be repeated, how much variability in the results might be expected? If the fitted LPA model is chaotic for a particular treatment, would it reliably be chaotic again? We deemed it crucial to obtain an estimate of how variable the estimated dynamic behaviors might be under stochastic repetitions of the experiment. This variability is sometimes termed “sampling variability,” though it must be kept in mind that what is “sampled” here are realizations of the stochastic population process.

We used a bootstrapping technique for time-series models. A version of the technique was used by Falck et al. (1995a, b); we incorporated a centering modification, described below, that was recommended in the statistics literature.

Recall that the residual vectors (Eq. 20) from the 24 populations were in two groups, control and treatment. For each group of residual vectors, the sample mean vector was calculated, and the resulting vector was subtracted from each residual in the group. These centered residuals were used in our bootstrapping procedure. The centering is theoretically important because the collection of centered residual vectors provides a statistically consistent estimate of the (possibly multivariate) noise distribution (Leger et al. 1992). In our data, the sample means of the residuals were near 0, and the centering was of little consequence.

To construct a bootstrapped time series for a single population, 40 residual vectors were sampled with replacement from the appropriate group (control or treatment) of centered residuals. The bootstrapped time series was started at the actual initial value $x_{0a} = x_0$, and was constructed with the relationship

$$x_{t}^a = \hat{h}(x_{t-1}^a) + \hat{\epsilon}_t^a$$

(29)

where $\hat{\epsilon}_t^a$ is the $t$th sampled residual vector, $\hat{h}(x_{t-1}^a)$ is the vector of functions in the model skeleton (Eq. 9) evaluated using the CLS parameter estimates and relevant values of $c_{ps}$ and $c_{pe}$. The resulting time series of vectors, $x_1^a, x_2^a, x_3^a, \ldots, x_m^a$, is a kind of simulated trajectory of life stages (square-root scale) for a popu-
lation in a given treatment under the estimated model. In this way, an entire bootstrapped data set of 24 populations in the form of the original data was constructed. The model was then refitted to the bootstrapped data set, yielding a set of bootstrapped CLS parameter estimates.

The process of generating a bootstrapped experiment (using the original CLS estimates) and calculating bootstrapped CLS estimates was repeated 2000 times, yielding 2000 sets of bootstrapped CLS parameter estimates. This collection forms a statistically consistent estimate of the multivariate sampling distribution of the CLS parameter estimates. The collection thus provides confidence intervals for parameters, or functions of parameters such as LEs. For each LPA model parameter, we used the 2.5th and 97.5th sample percentiles of the bootstrapped estimates as a 95% confidence interval.

Additionally, the collection provides estimates of how much variability there is in the types of dynamic behaviors estimated for each treatment. For each of the 2000 bootstrapped parameter sets, the dynamic properties of the corresponding LPA model were ascertained. First, the LEs and SLEs for each treatment were calculated with each bootstrapped parameter set. Second, if the LE was negative, the period of the attractor for the LPA skeleton was determined. The result for each treatment was a frequency count of different dynamic behaviors. At a given treatment, the proportion of the bootstrapped parameter sets that yielded, for instance, a two-cycle (i.e., a two-period cycle) is a valid estimate of the probability that a parameter set drawn from the CLS sampling distribution would produce a two-cycle.

RESULTS

Parameter estimates

The CLS (conditional least squares) estimates and 95% bootstrapped confidence intervals for the parameters in the stochastic LPA model (Eqs. 10–12) are given in Table 1. Since \( \mu_e \), the adult mortality probability, and \( c_{pa} \), the adult-pupa cannibalism coefficient, were fixed in the manipulated treatments, the point estimates and confidence intervals for these two parameters were obtained from the data on the controls. The confidence intervals indicate that the parameters are estimated with good precision; the half-widths of the confidence intervals ranged from around 2% to 11% of the CLS point estimates (Table 1). Also listed in Table 1 are the ML (maximum-likelihood) parameter estimates obtained from the Poisson/binomial model (Eqs. 4–8). The ML estimates were close to the CLS point estimates (Table 1). The ML estimates of \( b \), \( \mu_e \), \( c_{pa} \), and \( c_{ea} \) were well within the 95% CLS confidence intervals. The least similar ML estimate, that of \( c_{pa} \) in the control cultures, differed from the CLS estimate by only 10.5%. The binomial ML estimate for \( \mu_e \) in the control cultures was taken as fixed in the CLS estimation routines, and so is identical in both columns. Overall, the close similarity of the CLS and ML estimates suggests that the NLAR (nonlinear autoregressive) model (Eqs. 10–12) provides a good approximation to the Poisson/binomial model (Eqs. 4–8).

There was little or no correlation between the noise fluctuations of the state variables. The estimated covariances (Table 1) translate into the following correlations: \( \hat{\rho}_{12} = -0.10 \), \( \hat{\rho}_{13} = -0.096 \), and \( \hat{\rho}_{23} = -0.010 \) for the unmanipulated controls, and \( \hat{\rho}_{12} = -0.0030 \) for the manipulated treatments; despite the large sample sizes, none of these correlations are statistically different from 0. If the fluctuations were environmental in origin, we might expect that the departures of model and data would be related among the life stages (a good time interval for the L stage, say, would be related to whether or not the interval was good for the other stages). The lack of such correlation suggests that the noise is mostly demographic in nature.

The bootstrapped parameter sets provide additional information on the distributional properties of the parameter estimates. The quantile–quantile plots of Fig. 1 compare the observed quantiles of the standardized bootstrapped parameters to those of the standard normal distribution. The lack of departure from straight lines indicates that the estimates are well described with normal probability distributions. In Fig. 2 we show a scatterplot matrix of the bootstrapped parameter vectors. The histograms along the diagonal of the matrix also support the hypothesis that the parameter estimates are normally distributed. The lack of any strong correlations among the parameters, as indicated by the scatterplots, suggests that the model is not overparameterized. These results taken together suggest that the data provided good information about the model parameters.

| Table 1. Conditional least squares (CLS) and maximum-likelihood (ML) estimates of the LPA model parameters and their 95% bootstrapped confidence intervals. |
|---|---|---|---|
| Parameter\( ^\dagger \) | CLS estimate | ML estimate | 95% confidence interval |
| \( b \) | 10.45 | 10.67 | (10.04, 10.77) |
| \( \mu_e \) | 0.2000 | 0.1955 | (0.1931, 0.2068) |
| \( \mu_e \) | 0.007629 | 0.007629 | (0.006769, 0.008489) |
| \( c_{ea} \) | 0.01731 | 0.01647 | (0.01611, 0.01759) |
| \( c_{ea} \) | 0.01310 | 0.01313 | (0.01285, 0.01340) |
| \( c_{ea} \) | 0.004619 | 0.004135 | (0.004446, 0.004792) |

\( ^\dagger \) Definitions: \( b \) = number of larval recruits per adult per unit time in the absence of cannibalism; \( \mu_e \) = probability of larval mortality in one unit time; \( \mu_e \) = probability of adult mortality in one unit time; \( c_{ea} \) = larva–egg cannibalism coefficient; \( c_{pa} \) = adult–pupa cannibalism coefficient.
Estimated dynamics

The deterministic skeleton of the model (Eqs. 1–3), evaluated with the CLS point estimates, forecasts striking changes in dynamical behavior—from point equilibria, to cycles, to chaos—under our experimental conditions. Recall that manipulation of the adult mortality rate and adult recruitment rate provided the means of experimentally controlling the values of the parameters $m_a$ and $c_{pa}$. The estimated LPA skeleton yielded an estimate of dynamical behavior at each treatment value of $c_{pa}$ as well as an estimate of the LE (Table 2). A bifurcation diagram was computed with the CLS estimates to show how the estimated attractor changes as the parameter $c_{pa}$ changes (Fig. 3A). Fig. 3A and Table 2 together show the distinctive sequence of transitions in the estimated attractor of the model skeleton that occurs as the value of $c_{pa}$ is varied. The estimated bifurcation diagram (Fig. 3A) closely resembles the one reported earlier for this experiment using environmental noise (Costantino et al. 1997: Fig. 1); in fact, all the parameter estimates except for $b$ from the environmental-noise analysis lie within the confidence intervals reported here in Table 1.

For the control cultures, the estimated attractor is a stable point equilibrium (Table 2). For the $c_{pa} = 0.00$ treatment, the estimated behavior is aperiodic cycling on an invariant loop, which has an LE (Liapunov exponent) of 0 (Table 2, Fig. 3A). The loop actually consists of two tiny separate loops in phase space, with deterministic trajectories visiting each tiny loop in alternating time steps, and the result is that the trajectories resemble 2-period cycles. The estimated attractors for the $c_{pa} = 0.05$ and $c_{pa} = 0.35$ treatments are chaotic; the corresponding estimated LEs are positive, and the attractors are “strange.” The $c_{pa} = 0.10$ and $c_{pa} = 0.25$ treatments are stable long-period cycles that seem to occur in period-locked “windows” among more complex regions of the bifurcation diagram. The corresponding LEs are negative. The graph of the estimated LE as a function of $c_{pa}$ (Fig. 3B) shows great variability in the LE for values of $c_{pa}$ below 0.48 (approximately). The $c_{pa} = 0.50$ treatment produces a stable 3-period cycle, while the $c_{pa} = 1.00$ treatment gives a stable 6-period cycle. However the 6-cycle occurs just after a period-doubling bifurcation of the 3-cycle, and the 6-cycle trajectories resemble a 3-cycle in appearance. The LE estimates for $c_{pa} = 0.50$ and $c_{pa} = 1.00$ are negative.

The estimated SLEs (stochastic Liapunov exponents) differed substantially from the LEs (Table 2). Only the control and the $c_{pa} = 1.00$ treatments had negative SLEs; all the rest of the treatments had positive SLEs. The graph of the estimated SLE value as a function of $c_{pa}$ (Fig. 3C) is positive over a large interval of $c_{pa}$ values below 0.57 (approximately); by contrast the graph of the estimated LE values is jagged, irregular,

**Fig. 1.** Normal quantile–quantile plots of the (standardized) bootstrap parameter estimates. Units on both axes are standard deviations. A–E portray results for $b$, $m_l$, $c_{el}$, $c_{ea}$, and $c_{pa}$, respectively. See Table 1 for definitions of parameters.
and has negative as well as positive portions in that region (Fig. 3B).

The bootstrapped parameter estimates allow us to gain an idea of how variable are the estimates of dynamic behavior. The claim, for instance, that the attractor for \( c_{pa} = 0.10 \) is a 26-cycle (Table 2) is merely a "point estimate," that is, the claim relies on a point estimate of the model parameters. In fact, the 26-cycle is a narrow region of a complex bifurcation diagram (Fig. 3A), and the bifurcation diagram itself changes when model parameters change. If the point estimates were changed to other values within their confidence intervals (Table 2), the behavior of the skeleton at \( c_{pa} = 0.10 \) might be, say, a cycle with some other period.

Each of the 2000 bootstrapped parameter vectors provided an estimated LE, an estimated SLE, and an estimate of the dynamical behavior of the skeleton attractor for every treatment. The 2.5 and 97.5 percentiles of the 2000 bootstrapped LE and SLE values at each treatment were used to form 95% confidence intervals for the LE and SLE of that treatment (Table 2). The control and \( c_{pa} = 0.10, 0.25, 0.50, \) and 1.00 treatments,

### Table 2. Deterministic attractors and estimates (and 95% bootstrapped confidence intervals) of deterministic and stochastic Liapunov exponents (LE).

<table>
<thead>
<tr>
<th>Treatment†</th>
<th>Attractor</th>
<th>Deterministic LE</th>
<th>Stochastic LE</th>
</tr>
</thead>
<tbody>
<tr>
<td>Control</td>
<td>equilibrium</td>
<td>-0.046 (-0.047, -0.045)</td>
<td>-0.044 (-0.045, -0.044)</td>
</tr>
<tr>
<td>( c_{pa} = 0.00 )</td>
<td>invariant loop</td>
<td>0.000 (-0.036, 0.000)</td>
<td>0.009 (-0.007, 0.014)</td>
</tr>
<tr>
<td>( c_{pa} = 0.05 )</td>
<td>chaotic</td>
<td>0.029 (-0.227, 0.029)</td>
<td>0.045 (-0.042, 0.046)</td>
</tr>
<tr>
<td>( c_{pa} = 0.10 )</td>
<td>26-cycle</td>
<td>-0.109 (-0.115, -0.0004)</td>
<td>0.060 (0.056, 0.062)</td>
</tr>
<tr>
<td>( c_{pa} = 0.25 )</td>
<td>8-cycle</td>
<td>-0.076 (-0.090, -0.071)</td>
<td>0.068 (0.066, 0.071)</td>
</tr>
<tr>
<td>( c_{pa} = 0.35 )</td>
<td>chaotic</td>
<td>0.096 (-0.066, 0.100)</td>
<td>0.053 (0.049, 0.055)</td>
</tr>
<tr>
<td>( c_{pa} = 0.50 )</td>
<td>3-cycle</td>
<td>-0.089 (-0.101, -0.080)</td>
<td>0.019 (0.013, 0.025)</td>
</tr>
<tr>
<td>( c_{pa} = 1.00 )</td>
<td>6-cycle</td>
<td>-0.003 (-0.025, -0.0003)</td>
<td>-0.075 (-0.087, -0.063)</td>
</tr>
</tbody>
</table>

† The adult–pupa cannibalism coefficient is represented as \( c_{pa} \).
FIG. 3. Transitions in predicted deterministic dynamics for the point estimates of the parameters as reflected by (A) the bifurcation diagram for total population size (L-stage + P-stage + A-stage [see Table 3 for definitions of stages]), (B) deterministic Liapunov exponents (LE), and (C) stochastic Liapunov exponents, for \( c_{pa} \) values (adult–pupa cannibalism coefficients) ranging from 0 to 1. Arrows indicate experimentally fixed values of \( c_{pa} \).

had LE confidence intervals over values that were strictly negative. However, for \( c_{pa} = 0.10 \), most of the bootstrapped parameter vectors predicted high-period cycles with weak stability and LEs negative but near 0; the 97.5 percentile was just below 0 (Table 2). Similarly, for \( c_{pa} = 1.00 \), the 97.5 percentile was negative but close to 0. The \( c_{pa} = 0.00 \) treatment had an LE confidence interval with 0 included as the upper bound. This reflects an area of parameter space in which most parameter vectors yield stable cycles (LE < 0), but a measurable proportion of vectors result in invariant loops (LE = 0).

The LE confidence intervals for the \( c_{pa} = 0.05 \) and \( c_{pa} = 0.35 \) treatments include positive values. The LE confidence interval for the \( c_{pa} = 0.35 \) treatment, in fact, lies mostly in the positive range, with just a small portion overlapping into negative values. Later in this paper (see Bootstrapped attractors, below) we show that the negative values correspond mostly to high-period cycles of unusual periods (e.g., 19-cycles). Thus, we confidently claim that the attractor for the \( c_{pa} = 0.35 \) treatment is either chaotic or nearly aperiodic in relation to the length of our time series (40 time steps). The \( c_{pa} = 0.05 \) treatment is odd in that the LE point estimate is on the upper edge (to two decimal places) of the confidence interval. We show below (see Bootstrapped Liapunov exponents) how this stems from extreme skewness of the sampling distribution of LE estimates for the \( c_{pa} = 0.05 \) treatment.

The confidence intervals for the SLEs were more straightforward. Two of the treatments, the control and the \( c_{pa} = 1.00 \) treatments, had SLE confidence intervals that were strictly negative. Five of the treatments, \( c_{pa} = 0.05, 0.10, 0.25, 0.35, \) and 0.50, had SLE confidence intervals over strictly positive values. The \( c_{pa} = 0.00 \) treatment had an SLE confidence interval that included positive and negative portions. While one can debate about whatever property it is that SLE measures, estimating the SLE appears more stable than estimating the LE. We show below (see Bootstrapped Liapunov exponents) that the sampling distributions of the SLE estimates for all the treatments are fairly well behaved.

**Bootstrapped bifurcation diagrams**

The variability of the estimates of attractor behavior and deterministic LEs can be better understood by graphically exploring parameter space. The joint confidence intervals for the parameters can be regarded as a (hyper)rectangular region of parameter space, within which any parameter vector is plausible under the experimental evidence. (While the region is not strictly a joint 95% confidence region, the parameter estimates are nearly independent (see Fig. 2), and so the region forms an approximate (conservative) 95% confidence region.) The region generously indicates how much variability might be expected under repeated sampling. A fascinating picture of the variability in the estimates of the skeleton attractor resulted from plotting the \( c_{pa} \) bifurcation diagram for different parameter vectors in the region.

In Fig. 4 the \( c_{pa} \) bifurcation diagram is plotted for nine combinations of \( b \) and \( c_{el} \) values, with the other parameters held fixed at the CLS estimates. The three rows represent respectively the upper 95% confidence limit, the CLS estimate, and the lower 95% confidence limit, of the parameter \( b \). The columns represent the same limits for the parameter \( c_{el} \). The center diagram of Fig. 4 is, for reference, the CLS point estimate of the bifurcation diagram (as in Fig. 3). There is a strong qualitative similarity in the nine diagrams. For values of \( c_{pa} \) less than ~0.5 the diagrams show a region of complex behavior, with a large window of periodic
FIG. 4. Bifurcation diagrams for total population size (L-stage + P-stage + A-stage) plotted for all the combinations of the upper and lower 95% confidence limits of parameters $b$ (the number of larval recruits per adult per unit time in the absence of cannibalism) and $c_{el}$ (larva-egg cannibalism coefficient) for $c_{pa}$ values (adult–pupa cannibalism coefficients) ranging from 0 to 1.

behavior centered at approximately $c_{pa} = 0.2$. At a value of $c_{pa}$ just under 0.5, the complex behavior gives way abruptly to an extended region of stable 3-cycles. At particular values of $c_{pa}$, though, there is considerable local variation in the diagrams. For instance, at $c_{pa} = 1.00$, one of our treatment values, the lower row of diagrams (lower value of $b$) indicates that the attractor is estimated to be a 3-cycle, while in the upper two rows (increasing values of $b$), the estimated attractor has locally bifurcated into a 6-cycle. Similarly, around the treatment values of $c_{pa} = 0.05$ and 0.10 the bifurcation diagram is locally wispy; the dynamics of the attractors vary between cycles and chaos in intricate lacelike patterns.

The $c_{pa} = 0.35$ treatment lies within the dark, complex behavior region in all nine bifurcation diagrams (Fig. 4). The strange complexities of the attractors are revealed by zooming in for a close-up view. In Fig. 5 the interval from $c_{pa} = 0.34$ to 0.36 in the nine bifurcation diagrams is magnified. Between dark areas of aperiodic chaotic behavior, thin windows appear in which stable, high-period cycles undergo period-doubling bifurcations. Note the considerable variation in the locations of the windows. Similar features would be revealed, fractal-like, upon further magnification. By changing a parameter value a small amount in such regions, the behavior of the attractor can change considerably, from stable cycles to chaos, or the reverse. The parameter values giving rise to periodic behavior ($LE < 0$) in such regions are sets of positive measure (they have positive volume in parameter space), as are the parameter values giving rise to chaotic attractors ($LE > 0$).

**Bootstrapped Liapunov exponents**

Histograms of the bootstrapped LE values show how variability in parameter estimates becomes propagated into the LE estimates (Fig. 6). The bootstrapped LE values for the control treatment form a conventional bell-shaped histogram. The LE values represent estimated local stability strengths of the stable point equilibrium of the skeleton. The LE histograms for the $c_{pa} = 0.25$ and 0.5 treatments are also reasonably bell shaped. The LE values for these treatments represent the estimated local stability strengths of the cyclic attractors. The $c_{pa} = 0.25$ histogram actually extends into the region of positive LE values; 6 out of 2000 bootstrapped LEs were positive (not visible in the resolution of Fig. 6). The LE histograms for the other treatments
are highly skewed and peculiar (Fig. 6). For the \( c_{pa} = 0.00 \) treatment, most of the bootstrapped LE values are negative, but there is a spike of frequency at 0. The sampling distribution of the LE estimates here is apparently a mixed discrete and continuous distribution, with the positive probability at 0 giving the proportion of times the dynamic behavior is estimated to be aperiodic cycling on an invariant loop. The \( c_{pa} = 0.10 \) treatment has a left-skewed, ramp-shaped histogram of LE values in the negative region, with a visible little tail extending into the positive region (38 out of 2000 bootstrapped LEs were positive). The \( c_{pa} = 1.00 \) treatment has a left-skewed, J-shaped histogram of LE values. The LE values for \( c_{pa} = 1.00 \) were all strictly negative, though the histogram peak is near 0: the estimated cycles are weakly stable.

Though the LE histograms for the \( c_{pa} = 0.05 \) and 0.35 treatments have negative portions, they extend considerably into the positive region (Fig. 6). For the \( c_{pa} = 0.05 \) treatment, the skeleton attractor is highly variable in this region of parameter space (recall Fig. 4). A variety of stable cycles (1218 out of 2000), as well as chaos (782 out of 2000), are plausible behaviors estimated for the attractor, according to the bootstrapped LE values for \( c_{pa} = 0.05 \) (Fig. 6). The bulk (1670 out of 2000) of the bootstrapped LE values for the \( c_{pa} = 0.35 \) treatment are positive (Fig. 6). The remaining negative portion of the LE values for that treatment correspond to small windows of high-period stable cycles in an otherwise complex bifurcation diagram (recall Fig. 5).

The histograms for the bootstrapped SLE values were bell shaped for all the treatments (Fig. 7). The control and \( c_{pa} = 1.00 \) treatments had SLE histograms that were unambiguously within the negative region. The \( c_{pa} = 0.05, 0.10, 0.25, 0.35, \) and 0.5 treatments had SLE histograms that were unambiguously positive. The \( c_{pa} = 0.00 \) treatment had an SLE histogram that straddled 0. The normal-like shape of the histograms suggests that the SLE is a stable function of the model parameters for which the conventional asymptotic normality theory of maximum-likelihood estimates might apply (e.g., Lehmann 1983). We remark that the calculation of the bootstrapped SLEs for these histograms with appropriate numerical precision and safeguards (see Appendix) required weeks of running time on a contemporary Pentium computer.

**Bootstrapped attractors**

The 2000 bootstrapped parameter sets provided, for each treatment, 2000 estimated attractors. We summarized the frequencies of different dynamical behaviors of those attractors in a series of pie diagrams (Fig. 8). The control and \( c_{pa} = 0.50 \) treatments each showed...
entirely one type of behavior: stable point equilibrium for the control, and stable 3-cycles for the $c_{pa} = 0.50$ treatment. The $c_{pa} = 0.25$ treatment displayed stable 8-cycles nearly entirely, with a tiny portion (0.3%) of the attractors being chaotic. These three treatments had ordinary bell-shaped histograms of bootstrapped LE values (recall Fig. 6). The $c_{pa} = 1.00$ treatment was split roughly evenly between estimates of 3-cycles and 6-cycles (Fig. 8); the basic uncertainty arises because the value $c_{pa} = 1.00$ lies near a point where a 3-cycle bifurcates (recall Fig. 4).

Combinations of more complex behaviors were estimated for the $c_{pa} = 0.00, 0.05, 0.10, \text{and} 0.35$ treatments. At $c_{pa} = 0.00$, the spike of bootstrapped LE values at 0 in the histogram (Fig. 6) corresponds to the invariant loops (27%) indicated in the pie diagram (Fig. 8), while the negative LE values for that treatment (Fig. 6) were all stable 2-cycles (Fig. 8). Recall that the invariant loops in the “point estimate” of the attractor consisted of two tiny separate loops, producing trajectories resembling 2-cycles. For the $c_{pa} = 0.05$ treatment, 39.1% of the bootstrapped parameter sets gave chaotic attractors, and the remaining sets produced unusual stable cycles (Fig. 8). The $c_{pa} = 0.10$ treatment showed mostly stable cycles of unusual periods (e.g., 13-cycles). The $c_{pa} = 0.35$ treatment is our showcase for chaos. For that treatment, 83.5% of the bootstrapped parameter sets produced chaotic attractors, 7.1% produced stable 19-cycles, and the rest produced other stable cycles of higher periods (Fig. 8). In the unlikely
possibility that the behavior of the $c_{pa} = 0.35$ treatment is a stable cycle, it would probably have a high period relative to the length of the time series ($q = 40$) and occur in a small periodic window of parameter space within a larger region of chaos (Fig. 5).

Model evaluation

The generalized $R^2$ values for the experimental populations were very high (Table 3). The L-stage (feeding larvae) $R^2$ values were generally lower than those for the P-stage (pupae and near pupae). While three of the L-stage $R^2$ values were between 0.4 and 0.5, twenty of them were above 0.75, and twelve were above 0.9. For the P-stage, only two $R^2$ values were below 0.9; both were for control populations. The lowest $R^2$ value for the P-stage in all of the treatment populations was 0.96 ($c_{pa} = 0.00$, replicate 15). Many treatment $R^2$ values for the P-stage were above 0.99. The greater variability of the L-stage is easy to interpret in that recruits arise from the intrinsically variable processes of egg-laying and egg survival. P-stage recruits, by contrast, arise through a simpler survival process (see Eq. 2). We did not report $R^2$ values for the A-stage (sexually mature adults) in the treatment cultures because the experimental manipulations render the treatment A-stage populations essentially deterministic. Note that the control populations have L-stages and P-stages that are more variable (lower $R^2$ values, on average) than those in the treatments; the additional variability stems from the unmanipulated adults in the controls.

Residual diagnostics indicate that the stochastic portion of the stochastic LPA model (Eqs. 10–12) is a

Fig. 7. Histograms of the 2000 bootstrap estimates of the stochastic Liapunov exponents (SLEs) for each experimental treatment.
good representation of the noise in the data (Table 4). Significant first-order autocorrelation was detected in just 5 (10%) of the 51 time series for which there were residuals (L-, P-, and A-stages for 3 control populations, L- and P-stages for 21 treatment populations). Significant second-order autocorrelation was detected in just 7 (14%) of the time series; departure from a normal distribution was detected in 17 (33%) of the time series. These results are similar to results for 36 time series of this flour beetle strain (RR) reported for the first transitions experiment (3% first-order autocorrelated, 6% second-order autocorrelated, 33% non-normal; Dennis et al. 1997). That two thirds of the residual time series would fail to show evidence of departure from normality is remarkable, given that the series are long enough ($q = 40$) for the normality test to have reasonable power (Lin and Mudholkar 1980).

Various residual diagnostic plots, not reported here for space reasons, also showed excellent patterns (stable variances, normality). Note that the inferences we have made about parameters, dynamics, LE values, etc., are based on CLS estimation that makes no particular assumptions about the distributional form of the noise. The diagnostic tests and plots mainly suggest there are no major systematic or symptomatic patterns in the departures of model and data, allowing us to have greater confidence in the inferences made with the model.

That the estimated model skeleton accounts for a high degree of variability in the populations can be seen in time-series plots (Figs. 9 and 10). In Fig. 9, the L-, P-, and A-stage abundances are plotted for one representative population from the control and $c_{pa} = 0.00$, $0.05$, and $0.10$ treatments, along with the one-time-step forecasts of the fitted model. Fig. 10 gives the same plots for the $c_{pa} = 0.25$, $0.35$, $0.50$, and $1.00$ treatments. The patterns of changes in dynamics and variability from treatment to treatment are captured well by the model, from the stable point equilibrium of the control, to the irregular behavior of $c_{pa} = 0.35$, to the strong periodic signals in the $c_{pa} = 0.50$ and $1.00$ treatments. Occasionally the cyclic oscillations seemingly damp to a point equilibrium (for example, from $t = 8$ to $t = 16$ in the $c_{pa} = 0.10$ treatment, and from $t = 8$ to $19$ in the $c_{pa} = 1.00$ treatment); the model in fact predicts such behavior in the form of transient "flybys" of an unstable equilibrium lying on a stable manifold (Cushing et al. 1998b).

Phase-space plots and stochastic simulations

The agreement of model and data can be visualized better in phase space. In the left columns of Fig. 11...
Table 4. Residual analyses for each replicate. First-order ($\hat{\rho}_1$) and second-order ($\hat{\rho}_2$) sample autocorrelations and Lin-Mudholkar ($z$) test statistic for normality are reported.

<table>
<thead>
<tr>
<th>Treatment</th>
<th>Replicate no.</th>
<th>L-stage</th>
<th>P-stage</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>$\hat{\rho}_1$</td>
<td>$\hat{\rho}_2$</td>
<td>$z$</td>
</tr>
<tr>
<td>Control</td>
<td>4</td>
<td>0.47</td>
<td>0.27</td>
</tr>
<tr>
<td></td>
<td>11</td>
<td>0.51</td>
<td>0.43</td>
</tr>
<tr>
<td></td>
<td>24</td>
<td>0.66</td>
<td>0.43</td>
</tr>
<tr>
<td></td>
<td>12</td>
<td>-0.29</td>
<td>0.19</td>
</tr>
<tr>
<td></td>
<td>15</td>
<td>0.36</td>
<td>-0.12</td>
</tr>
<tr>
<td>$c_{ps} = 0.05$</td>
<td>1</td>
<td>0.11</td>
<td>0.35</td>
</tr>
<tr>
<td></td>
<td>7</td>
<td>-0.12</td>
<td>0.26</td>
</tr>
<tr>
<td></td>
<td>20</td>
<td>-0.24</td>
<td>0.20</td>
</tr>
<tr>
<td>$c_{ps} = 0.10$</td>
<td>6</td>
<td>-0.23</td>
<td>0.29</td>
</tr>
<tr>
<td></td>
<td>10</td>
<td>-0.24</td>
<td>0.41</td>
</tr>
<tr>
<td></td>
<td>16</td>
<td>-0.17</td>
<td>0.00</td>
</tr>
<tr>
<td>$c_{ps} = 0.25$</td>
<td>8</td>
<td>0.02</td>
<td>-0.24</td>
</tr>
<tr>
<td></td>
<td>17</td>
<td>-0.16</td>
<td>0.31</td>
</tr>
<tr>
<td></td>
<td>21</td>
<td>0.09</td>
<td>-0.05</td>
</tr>
<tr>
<td>$c_{ps} = 0.35$</td>
<td>13</td>
<td>0.01</td>
<td>0.11</td>
</tr>
<tr>
<td></td>
<td>22</td>
<td>-0.04</td>
<td>0.19</td>
</tr>
<tr>
<td>$c_{ps} = 0.50$</td>
<td>9</td>
<td>-0.18</td>
<td>-0.06</td>
</tr>
<tr>
<td></td>
<td>14</td>
<td>0.16</td>
<td>-0.14</td>
</tr>
<tr>
<td></td>
<td>19</td>
<td>-0.12</td>
<td>0.09</td>
</tr>
<tr>
<td>$c_{ps} = 1.00$</td>
<td>3</td>
<td>0.10</td>
<td>0.16</td>
</tr>
<tr>
<td></td>
<td>18</td>
<td>0.20</td>
<td>0.27</td>
</tr>
<tr>
<td></td>
<td>23</td>
<td>0.11</td>
<td>0.19</td>
</tr>
</tbody>
</table>

Notes: There is significant (0.05 level of probability) $j$th-order autocorrelation if $|\hat{\rho}_j|$ exceeds 0.31. There is significant (0.05 level of probability) departure from normality if $z > 1.96$. The diagnostic statistics for the A-stage residuals of the control treatments are: $\hat{\rho}_1 = 0.26$, $\hat{\rho}_2 = 0.23$, $z = -2.10$ for replicate 4, $\hat{\rho}_1 = -0.00$, $\hat{\rho}_2 = -0.02$, $z = 1.18$ for replicate 11; and $\hat{\rho}_1 = 0.14$, $\hat{\rho}_2 = 0.01$, $z = 2.11$ for replicate 24.

(A, C, E, G) and Fig. 12 (A, C, E, G), we plotted the data triples $[t, p, a]$ from each treatment in phase space after removing initial transient observations. Open circles are the data points; the dark circles or lines represent the estimated stable attractor for that treatment. The model attractors capture the transitions in dynamic behavior of the data from treatment to treatment extremely well. Note the tight cloud of points around the stable point equilibrium for the control treatment (Fig. 11A), the two clouds of points around the small invariant loops for the $c_{ps} = 0.00$ treatment (Fig. 11C), the irregular, triangular-shaped clouds of data points that accompany the three-pointed cyclic or chaotic attractor of the $c_{ps} = 0.05, 0.10, 0.25,$ and 0.35 treatments (Figs. 11E and G, 12A and C), and the clustering of points around the 3-cycle and 6-cycle of the $c_{ps} = 0.50$ and 1.00 treatments (Fig. 12E and G).

Clearly the data in Figs. 11 (A, C, E, G) and 12 (A, C, E, G) are influenced strongly by the deterministic forces present in the model. Also, there are clear departures of data from model. Does the variability predicted by the stochastic model resemble the variability in the data? The right columns of Figs. 11 and 12 (B, D, F, H) depict stochastic triples simulated using the estimated stochastic model (Eqs. 10–12, with parameter values given by Table 1), with initial transients removed. For the purpose of the simulations, the noise vector $E$, was assumed to have a multivariate normal distribution (recall that multivariate normality was not assumed for the estimation methods). The simulations are thus shown side by side with the corresponding data for each treatment. The resemblance of the left and right plots for each treatment is striking. The stochastic model produced cloud patterns of observations around the estimated attractors that can hardly be distinguished visually from the data. In ecology, such concordance of model and data across experimental manipulations is rare.

**DISCUSSION**

Asymptotic dynamics and stochasticity

Asymptotic dynamics (“final states”) such as equilibria, cycles, and chaos are deterministic mathematical notions defined in terms of equations and arbitrarily large temporal extents. Physical systems, being temporally finite, do not have asymptotic properties. Even so, these precise mathematical notions are useful for describing and understanding physical systems. (Indeed, the mathematical notions are perhaps nothing more than idealized abstractions gleaned from the observation of physical systems.) While a physical system cannot be at equilibrium, or on a limit cycle, or chaotic in a strict mathematical sense, these model concepts are useful as approximations to physical reality. If the state of a physical system is well described by a de-
terministic model, investigators commonly ascribe the mathematical dynamic of the model to the system. However, in population systems the role of chaos and other deterministic nonlinear dynamic behaviors, both asymptotic and transient, must be understood in the context of stochasticity.

Ecological populations are stochastic. Observed time series, from completely censused laboratory populations cultured under near-constant environmental conditions (Costantino and Desharnais 1981, Dennis and Costantino 1988) to extensive field surveys of population abundances (Woiwod and Hanski 1992), are characterized by pervasive, unexplained noise. In an analysis of 59 laboratory and field time-series data sets, Ellner and Turchin (1995) found that low-dimensional deterministic dynamics seldom accounted for more than half of the total variability in the data. While incorporation of weather covariates in time series (Roth-
Fig. 10. Time-series data (●) and one-step predictions from the fitted model (○) for representative cultures from four experimental treatments: $c_{oa} = 0.25, 0.35, 0.50$, and 1.00. The unit time is two weeks.

ery et al. 1997, Dennis and Otten 2000) will potentially account for some of the unexplained variability in field populations, population ecologists are a long way from being able to rely on “laws” of population dynamics in the form of differential or difference equations.

Noise in ecological systems has three major impacts on attempts to build appropriate mathematical models of population dynamics and to interpret population behavior in terms of model dynamic behavior. First, the noise must be included as an integral part of the model. Second, model parameters, and the resulting model dynamics, are estimated with statistical uncertainty. Third, the noise fundamentally alters the dynamic behaviors of the model. We discuss each of these impacts in turn, with reference to our results.

1. **Modeling the noise.**—If mathematical population models are to be built and tested as serious scientific hypotheses, the models must somehow be connected to data. Such connection implies that the noise must be modeled, in order to construct an appropriate estimating function for the model parameters (based on likelihood or conditional sums of squares, for instance).
The noise model component should be evaluated with data, using statistical diagnostic procedures. There is reason for encouragement; recent studies suggest that constructing useful stochastic models is feasible for various ecological systems. In at least some studies, diagnostic analyses of fitted models revealed noise with a simple probabilistic structure, such as a normal distribution (Dennis and Costantino 1988, Dennis et al. 1991, 1995, 1998, Kemp and Dennis 1993, Dennis and Taper 1994).

In the present study, the departures of data from the model skeleton were well described by a model of demographic variability. We hypothesized that the survival of a cohort of flour beetles through a life stage would resemble a simple binomial random process, and that recruitment of small larvae would be similar to a Poisson distribution. The binomial or Poisson-distribution parameters were modeled as nonlinear functions of the state variables in such a way that the mean trend for the system predicted over one time unit was the deterministic LPA model. The binomial and Poisson distributions had the property that the stochastic variability would appear as approximately normally distributed “noise” on the square-root scale. In the 24 experimental populations, the normal noise model provided overall an excellent description of the residual variation in the populations after the nonlinear effects predicted by the LPA model had been estimated and accounted for in the time series.

2. Estimating the dynamics.—When models are connected to data and parameters are appropriately estimated, interpretations of dynamic population behavior come with “confidence intervals.” In an earlier study, using a different strain of flour beetle, we displayed a joint parameter confidence region for unmanipulated control populations that contained stable 2-cycles as well as stable points (Dennis et al. 1995: Fig. 3). When a confidence region contains chaos, the situation can be far more complicated. Even though a point estimate of deterministic model parameters corresponds to a chaotic regime, a bewildering variety of other dynamic states could be embedded in the parameter confidence region. In fact, it was informative in the present study to divide the entire estimated sampling distribution of the parameter estimates into “confidence sets,” in which a degree of confidence is associated with a particular type of dynamic behavior. For instance, we are 39% confident that the $c_{pa} = 0.05$ treatment has a skeleton with a chaotic attractor, 23% confident that the attractor is a 5-cycle, and 38% confident that the attractor is cyclic with some other period (Fig. 8).

Based on our stochastic model and estimation procedures, we are 83.5% confident that the $c_{pa} = 0.35$ treatment has an underlying chaotic attractor, and 16.5% confident that the attractor is an unusual high-period cycle such as a 19-cycle (Fig. 8). The confidence sets provide strong support for our claim that the fluctuations of the $c_{pa} = 0.35$ treatment populations are largely explained by complex nonlinear behavior arising from simple, known population forces.

3. Emergent dynamic behavior.—Noise combines with deterministic forces to produce emergent dynamic
behaviors. Indeed, the mathematical notions of stability, periodicity, and chaos defined for deterministic systems do not strictly apply to stochastic systems. Instead of a chaotic stochastic system, or a periodic stochastic system, it is more appropriate to speak of the influence of a chaotic or periodic skeleton on the stochastic system. The skeleton and the noise together determine the dynamics (Leirs et al. 1997); the skeleton merely influences the dynamics.

The NLAR (nonlinear autoregressive) model (Eq. 9) can serve as a multivariate stochastic version of many types of ecological population models (Dennis et al. 1995, 1998), under demographic (square-root scale) or environmental (log scale) stochasticity. Such models have emergent dynamic properties that must temper discussions of nonlinear dynamics in ecology.

3a. The attractor of an NLAR model is a stationary probability distribution. The stationary distribution, if it exists, gives the long-run proportion of time that the system spends in a region of phase space as an integral of the stationary probability density function over that region. The population trajectories under Eq. 9 typically do not become confined to a reduced-dimension set, such as a point, cycle, or strange attractor. Rather, all regions of phase space are eventually visited. The stationary probability density function is typically smooth, instead of fractal like the invariant measure of a strange attractor, though the density might have many modes.

The stationary distribution for the stochastic LPA model (Eqs. 10–12) was estimated to undergo distinctive changes in response to our experimental manipulations of adult recruitment and mortality. The distributions under the different experimental treatments were manifested in model simulations as clouds of points around underlying deterministic attractors (Figs. 11 and 12). The resemblance of the model and data clouds, in particular the degree of variability around the attractors and the responses of the cloud patterns to the treatments, was striking (Figs. 11 and 12). Most of the probability in these stationary distributions was concentrated near the underlying attractors, suggesting that the attractors strongly influence the stochastic-model dynamics. Identifying the best statistical methods for comparing these fitted, exotic multivariate distributions with data is an open question that we are currently researching.

3b. Transient phenomena recur. Noise continually stirs stochastic systems, knocking trajectories away from the attractors of the skeleton. Transient phenomena, such as visitation to the neighborhood of an unstable equilibrium, eventually recur. Sometimes an unstable point (or other invariant sets such as an unstable cycle or fractal set) resides on a reduced-dimensional stable manifold, so that trajectories near the manifold tend to move toward the unstable point before moving away toward a stable attractor (Cushing et al. 1998b). The effect is for trajectories in the stochastic system to display occasional “fly-bys” of divergence regions of phase space, followed by fresh transient returns to the attracting region. As noted earlier (see Results: Model evaluation), recurring fly-bys of an unstable point equilibrium are evident in time series reported in the present study (Fig. 10). Long-run time spent near divergence regions is accounted for in the stationary
distribution. This recurring divergence implies a sensitivity to initial conditions of a kind fundamentally different from deterministic chaos (see Rand and Wilson 1991).

3c. Noise can cause transitions between multiple deterministic final states.—Stochasticity, in conjunction with the presence of unstable invariant separatrix sets and the nature, location, and size of basins of attraction, can “bump” model trajectories from one attracting deterministic state to another. For example, noise may cause a cycle to shift phase (Henson et al. 1998) or, in a regime with multiple deterministic attractors, may cause an orbit to jump from one cyclic attractor to another (Henson et al. 1999).

Influence of chaos on ecological populations

In a strict mathematical sense, populations cannot be chaotic for three reasons. First, even if populations were not stochastic, they are of finite temporal extent and thus do not have asymptotic dynamics such as equilibria or chaos. Second, given that populations are stochastic, the dynamics of their models must be estimated with confidence intervals, in which may be embedded dynamics other than chaos. Third, because of the interactions of stochasticity and deterministic influences, populations are not confined to a chaotic attractor so long as even a little stochasticity is present.

Instead, a more realistic hypothesis for a particular population might be that the population fluctuations are strongly influenced by a skeleton with, say, a strange attractor. Indeed, they might be strongly influenced by an unstable chaotic set through recurring transient behavior near the set. If so, then useful understandings and predictions might be accomplished with models that combine deterministic and stochastic aspects, and that is what we have demonstrated here.

Furthermore, an important take-home message of nonlinear dynamics for ecologists is not chaos per se, but rather that systems can undergo transitions among different types of behaviors in response to changing conditions, and that these transitions might be predicted with suitable stochastic models. Our experiments were designed around a specific sequence of transitions predicted by a promising biological model. A chaotic skeleton was predicted to be sandwiched between characteristic cyclic or stable-point skeletons, like books between bookends, along a range of values of a control parameter. The model predicted that the skeletons were influential enough that the transitions would be observed in experimental populations in response to manipulations of the control parameter value. The overall $R^2$ values indicated that the fitted LPA skeleton accounted for 89.9% of the L-stage (feeding larvae) variability and 98.5% of the P-stage (between feeding larvae and sexually mature adults; pupae) variability in the experimental populations (Table 3). The data plotted in phase space showed clear transitions in behavior and closely matched the predicted cycles on either side and the predicted strange attractor in the middle.

Are any of the populations in this experiment influenced by chaotic dynamics? On the basis of the point estimates of the Liapunov exponents (LEs) (see Table 2), the deterministic attractors for the $c_p$ (i.e., the adult pupa cannibalism coefficient) = 0.05 and 0.35 treatments are chaotic. There is additional supporting evidence. Histograms of the bootstrap estimates of the LEs revealed the occurrence of positive values for both treatments (Fig. 6), with the frequencies of positive values being 39.1% and 83.5% for the $c_p = 0.05$ and 0.35 populations, respectively (Fig. 8). Bootstrapped bifurcation diagrams placed both treatments in regions of complex dynamic behavior interwoven with chaos and high-period cycles (Fig. 4). The deterministic skeleton accounted for 92.5% of the variability in the (transformed) L-stage abundances, and 98.9% of the P-stage abundances, in the six cultures allotted to the $c_p = 0.05$ and 0.35 treatments, according to the generalized $R^2$ value calculated for those populations. We conclude that the $c_p = 0.05$ and 0.35 populations are strongly influenced by chaotic dynamics.

Sensitivity to initial conditions

Sensitivity to initial conditions is a hallmark of deterministic chaos. In a deterministic system, such sensitivity is a straightforward concept, and is widely measured with the dominant LE. The LE is the early divergence rate of two trajectories that differ infinitesimally in initial conditions, averaged over the system attractor (point, cycle, strange attractor, etc.). The average is taken with respect to the invariant measure of the attractor. The deterministic attractor is generally a set of greatly reduced dimension in phase space. Conditions on the attractor (where the LE is calculated) do not typify conditions elsewhere in phase space. The estimated LEs of the LPA model skeleton were positive at $c_p = 0.05$ and 0.35 (Table 2, Fig. 3B), indicating that for these $c_p$ values the skeleton alone produces chaotic behavior, that is, trajectories on the predicted strange attractor have the property (on average) of sensitivity to initial conditions.

Sensitivity to initial conditions must be measured in some other fashion, however, for a stochastic system. As is clear from both model simulations and data, stochastic trajectories at $c_p = 0.05$ and 0.35 do not spend much time actually on the attractors, but do behave as if influenced by the attractor (Figs. 11 and 12). Do such trajectories, in their recurrent visitations of transient areas of phase space, have the sensitivity property? It depends on how that sensitivity is defined.

The definition embodied in the “stochastic LE” (SLE) averages the trajectory divergence rate over the stationary distribution. The average can be calculated from the ergodic property by taking a time average of the divergence rate over a single long trajectory (see Appendix, and also Ellner and Turchin 1995). This def-
initiation, proposed by Crutchfield et al. (1982) and studied and advocated by McCaffrey et al. (1992), would seem a natural extension of the deterministic sensitivity concept to stochastic systems. Indeed, the SLEs calculated for \( c_{pa} = 0.05 \) and 0.35 (Table 2, Fig. 3C) indicate that trajectories of the predicted stochastic model for these \( c_{pa} \) treatments have, on average, the sensitivity property, so defined.

According to the full stochastic model, the \( c_{pa} = 0.00, 0.10, 0.25, \) and 0.50 treatments are also predicted to have the sensitivity property; the SLEs are positive. At first glance the situation might appear peculiar: the skeleton of the model for each of these \( c_{pa} \) values is decidedly nonchaotic, while the full stochastic model shows sensitivity to initial conditions. Is the noise somehow causing sensitivity to initial conditions?

The explanation lies in the properties of the stochastic NLAR model (Eq. 9). The stationary distribution accounts for the long-run time spent in all portions of phase space, including regions of phase space where initially close trajectories governed by the skeleton tend to diverge. In some situations a significant proportion of time can be spent by a stochastic system in a divergence region; if such a system is allowed to run for a long time and then stopped at random and observed, chances are fair by the ergodic property that the system will be in a divergence region. If a positive SLE were taken as a definition of chaos, then noise could “induce chaos” by continually knocking the system into phase-space regions that are transient and locally diverging according to the skeleton.

In fact, it is possible to construct a stochastic logistic/Ricker model in which the skeleton predicts a critically damped, globally stable point equilibrium, yet the SLE is positive (Deshamais et al. 1997). The noise in the model frequently places the population at low abundance levels, well below the stable point equilibrium of the skeleton. Because zero is an unstable equilibrium, trajectories near it diverge (growing approximately exponentially toward the stable equilibrium point). While such a system displays, on average, “sensitivity to initial conditions,” it is not clear that population ecologists would want to label such a system “chaotic.” After all, noisy perturbation around a stable equilibrium is a leading alternative hypothesis to chaos concerning the nature of fluctuations in natural populations (Pool 1989a, b).

Thus, defining chaos in terms of a positive SLE confounds both stochasticity and complex nonlinear dynamics; it classifies noisy systems as chaotic as well as systems under the influence of low-dimensional, nonlinear forces. Instead, we find studying and quantifying the influence of biologically based model skeletons useful for identifying the role of nonlinear dynamics in population ecology. All along, the mathematical chaos theorists provocatively suggested that population fluctuations might be largely deterministic, possibly caused by simple but unidentified nonlinear interactions (May and Oster 1976). In other words, it was hypothesized that if some underlying deterministic skeleton could be identified correctly, then the population fluctuations might be substantially explained and understood. Because of the presence of noise, ecological populations cannot be strictly chaotic; however, there remains the prospect that some ecological populations can be strongly influenced by underlying deterministic forces.

We have demonstrated that experimental Tribolium populations are strongly influenced by dynamics contained within a simple nonlinear skeleton. The predictions of the skeleton accounted for most of the variability of L-stage and P-stage numbers of 24 experimental populations. Furthermore, the skeleton forecasted substantially different dynamics depending on the experimental treatments, and these different dynamic transitions were displayed by the experimental populations. For the treatments with parameter \( c_{pa} \) experimentally set at 0.05 and 0.35, the estimated skeleton predicted chaotic dynamics. The skeleton accounted for most of the variability of L-stage and P-stage abundances displayed by the three populations at the \( c_{pa} = 0.05 \) and 0.35 treatments. The statistical patterns in the leftover noise were well described by a model of demographic stochastic forces.

**Concluding Remarks**

While there is no such thing as strict mathematical chaos—or even equilibria or cycles—in biological populations, ecologists can be encouraged that simple nonlinear models can help unlock substantial gains in understanding population systems. Keys to transforming nonlinear models from scientific caricatures to testable scientific hypotheses are: (a) incorporating the noise as well as the signal in biologically based models, (b) explicitly connecting models and data, (c) focusing on statistics as well as mathematics, (d) focusing on real systems and observable state variables, (e) rigorously evaluating model performance, and (f) effectively combining biology, mathematics, and statistics in interdisciplinary teamwork. As always, a well-designed experiment can pose a strong challenge to a model and help cut through a confusing fog of alternative possibilities. The Tribolium system has been studied and modeled in ecology for over 70 yr. We are just beginning to attain a mathematical understanding of its workings and a new appreciation of its possibilities.

**Acknowledgments**

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Here we describe our numerical methods for the computation of the deterministic and stochastic Liapunov exponents for the LPA model. The calculations center around a times series of triples \((L_n, P, A_*)\) and a matrix function \(J_t\) (Eq. 13). In the case of the Liapunov exponent (LE) the triples are a trajectory from the deterministic skeleton. In the case of the stochastic LE (SLE) the triples are a trajectory from the full stochastic model.

One problem with the direct use of Eq. 14 is that the matrix product may be unstable. A workaround is to "scale" the matrix product at each iteration. Let, \(s_1, s_2, \ldots, s_t\) represent a set of positive scalars and let \(S_t = J_t/s_t\). Then, from Eq. 14,

\[
\lambda_t = t^{-1} \ln |s_1 s_{t-1} \cdots s_t S_t S_{t-1} \cdots S_1| = t^{-1} \ln |S_t S_{t-1} \cdots S_1| + t^{-1} \sum_{i=1}^t \ln s_i. \tag{A.1}
\]

The "trick" is to choose the \(s_t\) scalars so that the matrix product remains stable. We used

\[
s_t = |J_t S_t S_{t-1} \cdots S_1| \tag{A.2}
\]

with \(s_1 = |J_1|\). With this definition, \(|JS_t S_{t-1} \cdots S_1| = 1\), and from Eq. A1 we have

\[
\lambda_t = t^{-1} \sum_{i=1}^t \ln s_i. \tag{A.3}
\]

The Liapunov exponent can now be computed as an arithmetic mean of a sequence of scalars. At each iteration the Jacobian \(J_t\) is computed from the values of \((L_n, P, A_*)\) and multiplied with the scaled matrix product \(S_t S_{t-1} \cdots S_1\), and the new scalar \(s_t\) is obtained as the norm of the matrix product \(J_t S_t S_{t-1} \cdots S_1\). The matrix product \(J_t S_t S_{t-1} \cdots S_1\) is divided by \(s_t\) to obtain a new scaled matrix product \(S_t S_{t-1} \cdots S_1\). Since, at each iteration, the norm of the scaled matrix product equals 1, the matrix product remains stable.

The problem of numerical roundoff error must also be handled carefully. At each iteration one must recompute the average of the first \(t\) scalars, and the average of the next scalar in the sequence. \(s_t\) can produce a large roundoff error. This loss of accuracy accumulates with each iteration.

A related problem is deciding on convergence. How large a value of \(t\) must one use in Eq. A3 to approximate Eq. 15 reasonably? Ideally, one would like to check for convergence within some error tolerance level at each iteration. However, as \(t\) gets large, adding another scalar to the mean does not have much effect on the mean. For example, the mean of \(t = 1000000\) scalars will not be much different from the mean of \(t = 1000001\) scalars, and it will look as if the mean \(\lambda\) has converged.

We addressed the roundoff problem by implementing an algorithm where the average Eq. A3 is built up from a series of averages of subsequences of scalars. For example, the average \(\lambda_{16}\) is obtained as the average of \((s_1 + s_2)/2\) and \((s_3 + s_4)/2\). The average of \(\lambda_{16}\) is obtained as the average of \(\lambda_{16}\) and average of \((s_5 + s_6)/2\) and \((s_7 + s_8)/2\). To get \(\lambda_{16}\), an average for \(t = 9, 10, \ldots, 16\) is computed in the same way \(\lambda_{16}\) was computed and the averages for \(t = 1, 2, \ldots, 8\) and \(t = 9, 10, \ldots, 16\) are averaged. This process is repeated to get estimates \(\lambda_2, \lambda_4, \lambda_8, \lambda_{16}, \lambda_{32}, \text{etc}\). At every step in the process, averages only occur between numbers computed from subsequences of the same length. All computations were done using double precision in the programming language C++ on a 32-bit operating system.

This algorithm also provides a conservative method for deciding on whether or not the average of the sequence has converged. The estimate of \(\lambda\) for \(t = 1, \ldots, 2^t\) is obtained by averaging the mean of the subsequence for \(t = 1, \ldots, 2^{t-1}\) and the mean of the subsequence for \(t = 2^{t-1} + 1, \ldots, 2^t\). To decide on convergence, these two "subsequence means" are compared. If the absolute value of their difference is less than a certain error tolerance \(e\), then it is decided that convergence has occurred.

We have found that one must choose \(e\) conservatively, especially for the SLEs. This is because one is computing \(\lambda\) from a single realization of the stochastic process. Although the process is ergodic, variation still occurs among SLEs estimated from finite sequences. We always repeat the estimation procedure using a different random-number sequence and compare estimates to make certain they agree.

Convergence occurs more quickly if one "tosses out" an initial subsequence of \(s_t's\) from the computation of the mean. This is because the scaled matrix product \(S_t S_{t-1} \cdots S_1\) tends to "settle down" as \(t\) gets large.

In the special case of a periodic attractor, the deterministic LE can be computed quickly and accurately without using the sequence-averaging method. Assume the deterministic process approaches a cycle with fixed period \(n\). For large, \(t\), \((L_n, P, A) = (A_{t+n}, P_{t+n}, A_{t+n})\). From Eq. 14 we can write, for \(t = mn\),

\[
\lambda_{mx} = (mn)^{-1} \ln |(J_m J_{m-1} \cdots J_{m-n+1}) S_t S_{t-1} \cdots S_1| \tag{A.4}
\]

The matrices \(C_1, C_2, \ldots, C_n\) represent the product of the Jacobian matrices taken in groups of \(n\). As the system converges onto its period-\(n\) cycle, the matrix \(C_n\) will approach a constant value \(C^*\), which is the product of the Jacobian matrices evaluated at each point on the cycle in the order in which they are visited. As \(m \to \infty\), \(t \to \infty\) and \(C_m \to C^*\). Therefore, for large values of \(m\), the matrix product \(C_1 C_{n-1} \cdots C_l\) will approach asymptotically the value \(\phi^* M\), where \(\phi^*\) is the largest modulus of the eigenvalues of \(C^*\) and \(M\) is a matrix of constants. From Eq. 15 we have

\[
\lambda = \lim_{m \to \infty} \lambda_{mx} = \lim_{m \to \infty} (mn)^{-1} \ln |\phi^* M| = \lim_{m \to \infty} (mn)^{-1} \ln |\phi| = n^{-1} \ln \phi. \tag{A.5}
\]

The LE can be computed directly from the eigenvalues of the matrix \(C^*\). The matrix \(C^*\) can be obtained by iterating the system until it converges to its stable \(n\)-cycle. Then the Jacobian matrices \(J_{n+1}, J_{n+2}, \ldots, J_{n+k}\) are computed and the product \(C^* = J_{n+1} J_{n+2} \cdots J_{n+k}\) is obtained. Next the eigenvalues of \(C^*\) are computed using one of the many numerical algorithms designed for this purpose. The LE is simply the logarithm of the largest modulus of the eigenvalues divided by the period.

In practice, we used the following procedure. We iterated the model 50000 times to remove transients. Then we computed the scaled Jacobian product and generated the \(s_t\) scalars. If the model was deterministic, we tested for a cycle of period \(n \leq 100\). If a cycle was found, the average of \(n\) of the \(s_t's\) was the estimate of \(\lambda\). If the model was stochastic or no cycle was detected, we tossed out the first 50000 of the values \(s_t's\) and used the "subsequence averaging" procedure described above on subsequent values of \(s_t\), to generate an estimate of \(\lambda\).

Estimates of the deterministic LEs were usually obtained quickly. Convergence was much slower for the stochastic LEs; for example, for \(e = 0.0001\), convergence sometimes took over a million iterations. But convergence did occur in a consistent and repeatable fashion.